A facile synthesis of pyrrolo-(di)-benzazocinones via an intramolecular *N*-acyliminium ion cyclisation†

Frank D. King,* Abil E. Aliev, Stephen Caddick, Derek A. Tocher and Denis Courtier-Murias

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A facile, moderate to high yielding synthesis of hexahydro-(di)-benzazocinones is described via an intramolecular N-acyliminium ion cyclisation. The iminium ion intermediates are formed from the readily available 4,4-diethoxybutyl amides with an excess of triflic acid. For electron-withdrawing substituents, better yields were obtained from the pre-formed 2-hydroxypyrrolidine amides. From NMR studies, at ambient temperatures the pyrrolo-benzazocin-3-ones exist as a slowly equilibrating mixture of two conformations.

Introduction

We have an on-going programme of work directed towards the development of new chemistry to allow the synthesis of privileged structures for the assembly of high quality screening libraries. We are interested in the synthesis of conformationally restricted aryl-substituted alkylamines, privileged structures which are found in CNS drugs,1-3 and particularly interested in 1,2,3,4tetrahydro-isoquinoline and its homologues, for which there have been numerous reports on their biological activity. 4-13 As an example of a new synthesis of the 1,2,3,4-tetrahydro-isoguinoline core, we recently described a synthesis of (±)-crispine A via an intramolecular cyclisation of a readily synthesized N-acyliminium ion.14 In this paper, we describe the unexpected isolation of a pyrrolo-dibenzazocin-3-one (3) from a pyrrolo-isoquinolone synthesis via an immimium ion cyclisation and the application of this methodology to related systems.

Results and discussion

Whilst investigating the scope of this tetrahydro-isoquinoline synthesis, we attempted the triflic acid-mediated cyclisation of compound 1, which was readily prepared from biphenyl-2-acetic

Department of Chemistry, University College London, 20 Gordon Street, London, WC1H 0AJ, U.K.. E-mail: f.d.king@ucl.ac.uk

acid.15 On treatment of 1 with triflic acid (10 equivs.) in CHCl₃ we isolated the 6-membered cyclisation product 2 (64% yield) as expected (Scheme 1), but in addition a second, isomeric product. ¹H and ¹³C NMR spectra provided evidence that the product was the dibenzazocine 3 (29% yield). The key difference between the ¹³C NMR spectra of 2 and 3 were the number of aromatic carbon signals, 10 signals for 2, 6 of which were C-H, and 12 signals for 3, 8 of which were C-H. The present observation is, as far as we can ascertain, the first example of an acyliminium ion cyclisation onto an unactivated aromatic to form an 8-membered ring. 16,17

Although benzo-fused 7-membered rings have been extensively exploited in drug discovery, for example tricyclic antidepressants, 18 benzodiazepines¹⁹ and as enzyme inhibitors,²⁰ there have only been a few examples of pharmacologically active benzo-fused 8membered rings described in the literature. 21-24 To date, hexahydrobenzazocines have been prepared either by ring expansion, 21,25-27 by ring-closing metathesis, 28,29 and by intramolecular cyclisation via either a Staudinger-aza-Wittig procedure²² or iminium ion cyclisations of sulfonamides.30-33 We therefore believed that this finding deserved further investigation.

Following the isolation of 3, we decided to investigate the synthesis of the simpler hexahydro-benzazocinone 5 from 4phenylbutyramide 4 (Scheme 2), prepared in a similar manner to 1 from 4-phenylbutyric acid. Cyclisation with triflic acid gave a good yield (70%) of a colourless oil, tentatively assigned as the hexahydrobenzazocinone 5 together with a small amount of 1tetralone (8% yield). The reaction was carried out at between 0.05– 0.1 M concentration demonstrating that no special procedures (e.g. high dilution) were required to mediate this ring formation.

Scheme 1 Synthesis of the tetrahydro-dibenzazocine 3. Reagents and conditions: a) (COCl)₂, CH₂Cl₂, DMF, b) H₂N(CH₂)₃CH(OEt)₂, EtNMe₂, c) CF₃SO₃H, CHCl₃, heat.

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Scheme 2 Synthesis of benzazocines 5 and 6. Reagents and conditions: a) (COCl)₂, CH₂Cl₂, DMF, b) H₂N(CH₂)₃CH(OEt)₂, EtNMe₂ (100%), c) CF₃SO₃H, CHCl₃, heat (70%), d) LiAlH₄, THF, heat (88%), e) picric acid.

Under these strongly acidic conditions, it is likely that the acyliminium ion is protonated on the carbonyl to form the dicationic superelectrophile, which undergoes the cyclisation.³⁴

At ambient temperatures, the ¹H and ¹³C NMR spectra of 5 in CDCl₃ showed broadened signals for the majority of the peaks, which was attributed to an intramolecular dynamic process with a rate intermediate in the NMR timescale at temperatures near 298 K. Relatively sharp ¹H and ¹³C NMR spectra were measured at 333 K, which were used for the structural confirmation of 5. Further variable-temperature ¹H and ¹³C measurements, including two-dimensional exchange-correlated spectroscopy, 35 revealed the presence of two conformations in the dynamic process. In particular, two sets of peaks were observed at 213 K in the ¹H and ¹³C NMR spectra of 5 with the integral intensity ratio of 2.7:1. Proton H-2, the bridgehead proton of the major conformer, appeared as a doublet of doublets (J = 6.4 and 9.3 Hz), whereas that of the minor conformer showed only a doublet (J = 7.0 Hz). Based upon these values and the boundary values of the corresponding ${}^{3}J_{\mathrm{HH}}$ couplings predicted for L-prolines, 36 the five-membered ring of the major conformer is in the C^4 -exo conformation, whereas that of the minor comformer is in the C4-endo conformation. Here we use the previously suggested notation for L-proline conformations, ^{36,37} where the endo-/exo-orientation of the C4 atom is determined relative to the C^2 - C^1 bond. Overall, the $^3J_{\rm HH}$ couplings of both the five- and eight-membered rings were in favour of a two-site conformational equilibrium shown in Fig. 1, where the C⁴-exo-C⁸-endo form is the preferred major conformer. The free energy barrier for the conformational change from the C4-exo-C8-endo conformer into the C4-endo-C8-exo conformer for 5 was estimated to be 55 ± 1 kJ mol⁻¹ at the coalescence temperature of 266 K. For the reverse interconversion from the C4-endo-C8-exo conformer into the C4-exo-C8-endo conformer the free energy barrier was estimated to be $53 \pm 1 \text{ kJ mol}^{-1}$ at 266 K.

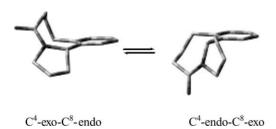


Fig. 1 A representation of the two conformers of 5 from NMR analysis.

In order to obtain a solid derivative to unambiguously confirm the molecular structure, 5 was reduced to the amine with LAH and a solid picrate salt 6 formed. Unfortunately good quality crystals of the picrate could not be obtained. However, the structure of this salt was confirmed by a low resolution X-ray crystal structure analysis. In addition, the structure of the corresponding free base was also confirmed by NMR measurements in CDCl₃ (Fig. 2).

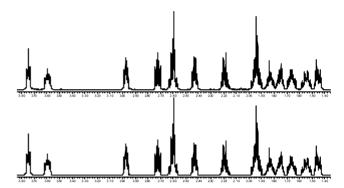


Fig. 2 Experimental (top, in CDCl₃, 333 K) and calculated (bottom) 600 MHz ¹H NMR spectra of the free base of 6. Fifteen protons of the five- and eight-membered aliphatic rings were included in the iterative fittings of the experimental spectrum.

The scope of this method was then investigated with respect to aromatic substituents, and the results are shown in Table 1.

As expected of an electrophilic cyclisation, electron-donating substituents increased the rate of reaction and gave higher yields, as exemplified by the p-tolyl and 3,4-dimethoxy analogues (5a and **5b** respectively). The 2,5-dimethoxy **5c** gave a much lower yield, though the reaction was still fast and could be run at ambient temperatures. Electron-withdrawing substituents both slowed the reaction and gave lower yields. Thus the 4-Br 4e and 4-phenyl 4f gave only poor yields of the benzazocinones 5e and 5f respectively. None of the desired product could be detected from the 4-Cl 4d cyclisation. In all cases, where a poor yield was obtained a significant quantity of insoluble, presumably polymeric material, was produced. For the very electron-rich 4b, the cyclisation could be achieved using 10 equivs. of TFA (1 h reflux, 90% yield).

In the ¹H and ¹³C NMR spectra of **5b** and **5e** at 213 K, again two sets of peaks were observed with the integral intensity ratios of 4.3:1 and 3.4:1 respectively. Similar to 5, the free energy barrier for the conformational change from the C4-exo-C8-endo conformer into the C⁴-endo-C⁸-exo conformer for **5e** was estimated to be $56 \pm$ 1 kJ mol⁻¹ at the coalescence temperature of 266 K. For the reverse interconversion from the C4-endo-C8-exo conformer into the

Table 1 Synthesis of substitued hexahydrobenzazocin-3-ones 5a-f

Cpd. no.	R	$T(h)^a$	Cpd. no.	R	Isolated yield (%)
4a	4-Me	4	5a	14-Me	75
4b	3,4-diMeO	1	5b	13,14-diMeO	80
4c	2,5-diMeO	21^{b}	5c	12,15-diMeO	26
4d	4-C1	18	5d	14-Cl	0
4e	4-Br	18	5e	14-Br	17
4f	4-Ph	0.5	5f	14-Ph	25

^a Time of reaction in CHCl₃ heated under reflux. ^b At ambient temperatures.

C⁴-exo-C⁸-endo conformer the free energy barrier was estimated to be $53 \pm 1 \text{ kJ mol}^{-1}$ at 266 K (Fig. 3).

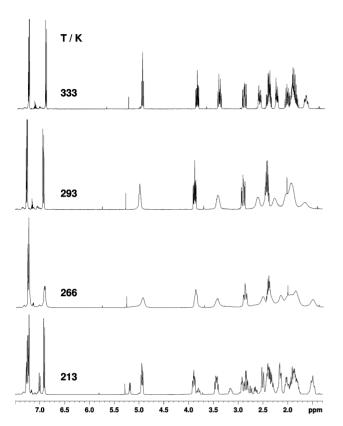


Fig. 3 The temperature dependence of the ¹H NMR spectra of 5e (CDCl₃, 400 MHz).

A sample of 5c was submitted for X-ray analysis, which confirmed the structure as the benzazocin-3-one (Fig. 4). Unlike the other benzazocinones, 5c showed sharp ¹H and ¹³C NMR spectra in CDCl₃ at 298 K with the ³J_{HH}-couplings indicating of a single C4-exo-C8-endo conformer, in agreement with the solid-state structure. Presumably the steric constraints of the 12and 15-substituents restricts the conformational freedom of the 8,5 system.

In our previous paper,14 a higher yield of tetrahydro-isoquinolinone was obtained from the N-acyl-2-hydroxy-pyrrolidine,

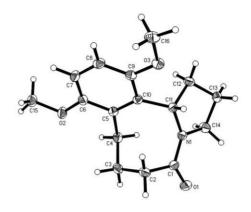


Fig. 4 Structure of hexahydrobenzazocinone 5c as determined by X-ray crystallography. ORTEP diagram (50% probability ellipsoids) showing the crystallographic atom-numbering scheme.

readily prepared by mild acid hydrolysis of the amidoacetal. Thus treatement of an acetone solution 4d-h with 1 M HCl solution for 15 min gave the intermediate N-acyl- 2-hydroxypyrrolidines **7d-h** in essentially quantitative yields (Scheme 3).

Previous experience had shown that N-acyl-2-hydroxy-pyrrolidines were too unstable to purify by column chromatography and so 7d-i were used without further purification. Triflic acid cyclisation of 7d-g gave higher yields of benzazocines 5d-g, than previously obtained from the amides (yields quoted are the overall yields from 4d-g). Cyclisation of the m-Br congener 7g gave almost exclusively the 13-Br product 5g, with only a trace of a less polar isomer, presumably the 15-Br, detectable by MS. Cyclisation of the 4-methoxy lactam 7i gave only a trace (TLC, MS) of the hexahydrobenzazocine, but an improved yield was obtained from the 3-Br, 4-MeO lactam 7h as a single isomer. A similar low yield was reported for the TiCl₄-mediated cyclisation of 1-(4methoxyphenylacetyl)-2-methoxypyrrolidine to the pyrrolidinotetrahydroisoguinoline.38

This methodology also works well for phenyl butyramides substituted in the alkyl chain. Cyclisation of the 4,4-diphenylbutyric acid amide gave a good yield of the phenyl-substituted benzazocinone, which from NMR, the isolated product was a mixture of isomers in a ratio of 20:1 (Scheme 4). The major

Scheme 3 Synthesis of the hexahydrobenzazocines 5d-i via the 2-hydroxypyrolidines 7d-i. Reagents and conditions: a) HCl, acetone, b) CF₃SO₃H, CHCl₃, heat.

Scheme 4 Synthesis of benzazocine 8; Reagents and conditions: a) (COCl)₂, CH₂Cl₂, DMF, b) H₂N(CH₂)₃CH(OEt)₂, EtNMe₂ (100%), c) CF₃SO₃H, CHCl₃, heat (73%).

isomer 8 was isolated as a solid and its structure confirmed by X-ray analysis (Fig. 5), which shows that the phenyl group is in a pseudo-equatorial orientation 'cis' to the bridgehead C4 proton.

Fig. 5 Structure of hexahydro-benzazocinone 8 as determined by X-ray crystallography. ORTEP diagram (50% probability ellipsoids) showing the crystallographic atom-numbering scheme.

Both the attempted application of this methodology to the synthesis of the 9-membered ring homologue from the 5phenylpentanoic acid amide 9 and the attempted cyclisation of the 4-phenylbutanoic acid amide of 5-aminopentananal diethylacetal 10 failed to give any cyclised products.

$$Ph(CH_2)_4CONH(CH_2)_3CH(OEt)_2 \ \ \boldsymbol{9}$$

Ph(CH₂)₃CONH(CH₂)₄CH(OEt)₂ 10

An alternative system was also evaluated where there is no possibility for the formation of a 6-membered ring (Scheme 5). The commercial α-phenyl-o-toluic acid was converted into the 4-aminobutyraldehyde diethylacetal amide derivative via the acid chloride. Cyclisation with triflic acid afforded the tetracycle 12a in an excellent yield. Both the ¹H and ¹³C NMR spectra were consistent with the proposed structure. The structure of 12a was confirmed by an X-ray structural analysis (Fig. 6). Cyclisation of the ortho-phenoxyamide derived from 11b also gave the cyclic product 12b, though in a slightly lower yield.

Although it is well known that acyliminium ions are more electrophilic than iminium ions,39 for comparison the Pictet-Spengler type cyclisation of the amine 13, prepared from 4 by reduction with LAH, to 6 was investigated.

$$Ph(CH_2)_4NH(CH_2)_3CH(OEt)_2$$
 13

Scheme 5 Synthesis of dibenzoazocines 12a and 12b: Reagents and conditions: a) (COCl)2, CH2Cl2, DMF, b) H2N(CH2)3CH(OEt)2, EtNMe2 (100%), c) CF₃SO₃H, CHCl₃, heat.

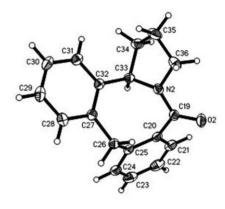


Fig. 6 Structure of hexahydro-dibenzazocinone 12a as determined by X-ray crystallography. ORTEP diagram (50% probability ellipsoids) showing the crystallographic atom-numbering scheme. There are two independent molecules in the asymmetric unit and only one of these is shown in the figure. The asymmetric unit was selected so that both molecules have the same R-stereochemistry.

No formation of 6 was observed under the acidic conditions of TFA or triflic acid 2 h in refluxing CHCl₃, or 2 M aqueous HCl at reflux for 24 h.

Conclusion

In conclusion, we have shown that pyrrolo-benzazocin-3-ones can be readily prepared from the commercially available 4aminobutyraldehyde diethyl acetal and the appropriate carboxylic acids in moderate to high yield via an intramolecular N-acyl iminium ion cyclisation. For electron-withdrawing substituents, pre-formation of the 2-hydroxy-pyrrolidine amides gave better yields. Two series of pyrrolo-dibenzazocines were also prepared. This method is not applicable for the synthesis of the 3-azabenzocyclononanes or the piperidino-benzazocines, nor was the Pictet-Spengler cyclisation successful. Our future work will concentrate on further investigation of the scope of this methodology and on the synthesis of 8-membered analogues of biologically active tetrahydroisoquinolines and benzazepines, and it is our belief that this chemistry will further open up the possibility of 8-membered rings for exploitation in, for example, chemical library synthesis and drug discovery.

Experimental

All reagents were commercially available, unless specified, and used without purification. The chloroform used was stabilised with amylene. All non-crystalline final compounds were found to be >95% pure by HPLC, and all crystalline compounds >98% pure. Solution ¹H and ¹³C NMR spectra (Tables 2, 3 and 4) were recorded on Bruker NMR spectrometers AMX300, Avance III 400, DRX500 and Avance III 600 equipped with z-gradient facilities. ¹H and ¹³C chemical shifts are given relative to TMS. Unless otherwise specified, spectra were recorded at 298 K. The ¹H spectra of **5b**, **5c** and **6** were analysed using full lineshape analysis.⁴⁰ Low temperature NMR measurements were carried out for hexahydro-benzazocinones 5, 5b and 5e. The value of the free energy of activation for 5 and 5e was calculated using the procedure described previously.⁴¹

4-(3-Bromophenyl)-butyric acid

4-(3-Bromophenyl)-4-oxo-butyric acid⁴² (4.8 g) and KOH (3.6 g) was dissolved in 35 ml ethylene glycol and hydrazine hydrate (2.4 ml) added. The reaction mixture was heated to reflux (140 °C) for 2 h, then heated to 220 °C (heating block temperature) to distill out the H₂O and excess hydrazine hydrate. After all the distillation had ceased, the reaction mixture was heated for 1 h, the cooled to room temperature, H₂O added (100 ml) and conc. HCl to acidic, then ice (50 g). On scratching with a glass rod, a solid had formed which was collected, dissolved in Et₂O (150 ml) and the product extracted into 1 M NaOH (2 × 50 ml). The aqueous solution was acidified with 2 M HCl and the product extracted

into CH_2Cl_2 (3 × 50 ml). The organic extracts were dried (MgSO4) and filter through a SiO₂ bed (~20 g), washing with Et₂O until no more non-polar material was extracted. The combined washings were evaporated in vacuo to give 3.1 g of pale yellow oil. ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3)$: $\delta 1.95 \text{ (quintet, 2H, } J = 7 \text{ Hz}), 2.39 \text{ (t, 2H, } J =$ 7 Hz), 2.65 (t, 2H, J = 7 Hz), 7.10–7.21 (m, 2H), 7.31–7.40 (m, 2H) including 7.35, d, 1H, J = 1.5 Hz). ¹³C NMR and DEPT (75 MHz, $CDCl_3$) $\delta = 25.6$ (CH_2), 33.3 (CH_2), 34.6 (CH_2), 122.5 (C), 127.2 (CH), 129.3 (CH), 130.0 (CH), 133.6 (CH), 143.6 (C), 180.0 (C).

Synthesis of 4-(3-bromo-4-methoxyphenyl)-butyric acid

A solution bromine (0.52 ml, 10 mmol) in CHCl₃ (10 ml) was added, drop-wise to a stirred solution of 4-(4-methoxyphenyl)butyric acid (1.94 g, 10 mmol) in CHCl₃ (30 ml) at ambient temperature and the stirring was continued for 1 h. The solvent was removed in vacuo and the residue triturated purified by column chromatography on silica, eluting with CH₂Cl₂-10% Et₂O/CH₂Cl₂. and triturated with petrol to give 1.4 g of white solid (50%). Mpt = 78–80 °C; ¹H NMR (300 MHz, CDCl₃): δ 1.95 (quintet, 2H, J = 7 Hz), 2.39 (t, 2H, J = 7 Hz), 2.65 (t, 2H, J = 7 Hz), 7.10–7.21 (m, 2H), 7.31–7.40 (m, 2H including 7.35, d, 1H, J = 1.5 Hz). ¹³C NMR and DEPT (75 MHz, CDCl₃) $\delta =$ 6.2 (CH2), 33.2 (CH2), 33.7 (CH2), 56.3 (CH3), 111.6 (C), 112.0 (CH), 128.4 (CH), 133.3 (CH), 134.9 (C), 154.3 (C), 180.0 (C).

General procedure for the synthesis of the amides

Oxalyl chloride (10 mmol) was added to a stirred solution of the appropriate acid (10 mmol) in DCM (30 mL) and 1-2 drops of

Table 2 ¹H NMR chemical shifts (δ_H, ppm) of hexahydrobenzazocinones in CDCl₃. The spectrum of 5c was measured at 298 K, all other spectra were measured at 333 K. The structure shows the proton and carbon numbering used in the NMR assignments in Tables 2-4. The relative orientations of the protons are defined relative to proton H-2c: "c" and "t" are used to denote protons with the cis and trans orientations relative to proton H-2c

Proton	5	5a	5b	5c	5d	5e	5f	6^a
2c	5.03	4.98	4.967	5.182	5.00	4.98	5.13	3.746
3c	2.40	2.40	2.412	2.520	2.44	2.42	2.49	2.192
3t	2.07	2.06	2.081	1.719	2.07	2.06	2.17	1.949
4c	1.87	1.86	1.888	1.813	1.90	1.89	1.92	1.832
4t	1.91	1.91	1.960	1.853	1.96	1.95	1.98	1.933
5c	3.87	3.87	3.863	3.838	3.90	3.88	3.94	2.430
5t	3.43	3.44	3.447	3.549	3.44	3.42	3.50	2.974
8c	2.49	2.48	2.508	2.371	2.47	2.44	2.57	1.546
8t	2.26	2.24	2.265	2.106	2.29	2.27	2.32	1.453
9c	1.70	1.66	1.713	1.447	1.70	1.68	1.77	1.662
9t	1.95	1.95	1.935	1.927	1.96	1.95	2.04	1.754
10c	2.66	2.48	2.584	3.207	2.64	2.62	2.73	2.722
10t	2.96	2.92	2.912	2.529	2.95	2.92	3.03	3.593
12	7.05	6.93	6.553	3.747 (OMe)	7.00	6.93	7.15	7.042
13	7.16	6.97	3.833 (OMe)	6.738	7.14	7.27	7.41	7.15
14	7.14	2.29 (Me)	3.836 (OMe)	6.689	_	_	_	7.14
15	7.12	6.92	6.638	3.747 (OMe)	7.13	7.26	7.36	7.13

" 2.590 ppm (H-7c) and 2.611 ppm (H-7t).

Table 3 Proton *J*-couplings (in Hz) of hexahydrobenzazocinones 5–5f and 6 in CDCl₃. The spectrum of 5c was measured at 298 K, all other spectra were measured at 333 K

Protons	5	5a	5b	5c	5d	5e	5f	6
2c-3c	7.1	7.1	6.99	6.15	7.1	7.1	7.1	7.55
2c-3t	7.1	7.1	6.98	9.69	7.1	7.1	7.1	7.74
3c-3t	-12.7	-12.7	-12.80	-12.57	-12.7	-12.9	-12.7	-12.87
3c-4t	5.1	5.0	5.29	3.29	5.2	5.1	5.0	5.29
3c-4c	7.6	7.5	7.58	6.51	7.5	7.4	7.5	10.06
3t-4c	8.7	8.7	8.41	10.81	8.5	8.7	8.6	5.83
3t-4t	7.3	7.2	7.25	6.68	7.0	7.1	7.0	10.72
4c-4t	-12.6	-12.5	-12.63	-12.41	-12.7	-12.6	-12.6	-12.54
4c-5c	8.6	8.5	8.42	8.65	8.4	8.4	8.5	8.63
4c-5t	8.1	8.1	8.12	9.06	7.9	8.2	8.3	3.74
4t-5c	4.1	4.2	4.41	3.44	4.3	4.2	4.3	7.68
4t-5t	8.1	8.1	7.85	7.45	7.9	8.0	8.1	8.38
5c-5t	-12.2	-12.2	-12.17	-12.21	-12.2	-12.1	-12.2	-9.14
8c-8t	-12.4	-12.3	-12.32	-11.88	-12.5	-12.5	-12.4	-15.00
8c–9c	4.8	4.9	4.79	5.38	4.8	4.7	4.9	11.97
8c–9t	12.8	12.8	12.72	13.27	12.7	12.8	12.8	3.34
8t–9c	4.2	4.1	4.03	2.61	4.1	4.0	4.1	3.35
8t-9t	4.2	4.0	4.18	4.47	4.1	3.9	4.1	5.37
9c–9t	-13.0	-12.8	-13.41	-13.06	-12.8	-12.8	-12.8	-13.62
9c-10c	4.0	4.9	3.71	3.90	3.8	3.9	3.9	6.71
9c-10t	11.7	11.9	11.82	13.55	11.8	11.9	11.9	5.31
9t-10c	5.0	4.9	5.04	3.04	5.0	5.0	4.7	5.32
9t-10t	3.9	3.9	3.80	4.09	3.9	3.9	3.9	8.85
10c-10t	-13.8	-13.8	-13.96	-13.29	-13.9	-13.9	-13.8	-12.82
Other	_	7.8 (12–13),	_	8.94 (13-14)	~8 (12–13),	~8 (12–13),	7.9 (12–13),	-12.57 (7c-7t), 3.09 (7c-8c),
		1.7 (13,15)		, ,	2.2 (13,15)	2.1 (13,15)	2.0 (13,15)	5.27 (7c–8t), 0.85 (7c–9t), 11.28 (7t–8c), 3.13 (7t–8t)

Table 4 $^{-13}$ C NMR chemical shifts (δ_C , ppm) of hexahydrobenzazocinones **5–5f** and **6** in CDCl₃. The spectrum of **5c** was measured at 298 K, all other spectra were measured at 333 K. The values of $^{1}J_{CH}$ couplings (in Hz) are also included in brackets for **5c**

Carbon	5	5a	5b	5c	5d	5e	5f	6
1-C _q	140.54	140.35	132.75	130.09	142.38	142.85	140.90	141.08
2-CH	62.80	62.93	62.53	59.44 (145)	62.35	62.40	63.11	65.65
$3-CH_2$	36.83	37.12	36.91	36.09 (134)	36.78	36.99	37.33	34.16
4-CH ₂	22.75	22.77	22.80	22.50 (131)	22.66	22.77	22.92	22.71
5-CH ₂	45.94	46.01	46.04	46.81 (143)	45.93	46.08	46.15	55.30
7	172.48	172.50	172.61	172.71	172.22	172.32	172.65	54.01
8-CH ₂	33.81	33.67	33.91	32.26 (131)	33.59	33.63	33.80	24.67
9-CH ₂	27.37	27.60	27.61	25.74 (132)	27.22	27.25	27.52	30.17
10-CH ₂	32.62	32.03	32.22	21.01 (131)	31.93	32.06	32.32	32.98
11-C ₀	138.16	135.01	130.96	127.48	136.60	137.18	137.25	140.19
12	131.49	131.46	115.20	151.02	132.81	133.22	132.17	130.50
13	126.57	128.31	148.66	109.47 (159)	127.58	130.67	126.38	125.92
14	127.61	135.97	147.89	108.31 (159)	132.29	120.29	141.04	126.86
15	126.90	127.63	111.61	150.19	126.91	129.95	125.84	127.13
Other	_	20.97 (14-Me)	56.54 (13-OMe), 56.24 (14-OMe)	56.21(143) (12-OMe), 55.75 (143) (15-OMe)	_	_	139.78 (<i>i</i> -Ph), 127.13 (<i>o</i> -Ph), 128.96 (<i>m</i> -Ph), 127.50 (<i>p</i> -Ph)	

DMF were added. After stirring at ambient temperatures for 2 h, by which time gas evolution had ceased, the solvent was removed by rotary evaporation *in vacuo*. The crude acid chloride was redissolved in DCM (20 mL) and was added, dropwise, to a stirred, cooled (0 °C) solution of 4-aminobutyraldehyde diethyl acetal (10 mmol) and Et₂NMe (12 mmol) in Et₂O (50 mL) over 5 min. The stirred reaction mixture was warmed to room temperature over 1 h. A 1 M aqueous solution of NaHCO₃ (30 mL) was then added and the mixture transferred to s separating funnel, shaken then allowed to settle. The lower aqueous layer was separated and the organic layer washed with brine (30 mL). The organic layer was collected, dried (K_2CO_3), filtered and the filtrate concentrated

in vacuo, then dried under high vacuum to give the essentially pure amides (>98% by HPLC) which were used without further purification.

2-Biphenyl-2-yl-*N***-(4,4-diethoxybutyl)acetamide (1).** Isolated as an oil (95% yield). ¹H NMR (300 MHz, CDCl₃): δ 1.17 (t, 6H, J=7 Hz), 1.40–1.56 (m, 4H), 3.16 (dt, 2H, J=7, 13 Hz), 3,37–3.66 (m, 4H), 3.52 (s, 2H), 4.42 (t, 1H, J=5 Hz), 5.35 (brs, 1H), 7.24–7.48 (m, 9H): ¹³C NMR and DEPT (75 MHz, CDCl₃) δ = 15.3 (CH₃), 24.6 (CH₂), 30.8 (CH₂), 39.3 (CH₂), 41.4 (CH₂), 61.3 (CH₂), 102.5 (CH), 127.3 (CH), 127.4 (CH), 128.0 (CH),

129.1 (CH), 130.5 (CH), 130.7 (CH), 132.5 (C), 140.9 (C), 142.6 (C), 170.9 (C).

N-(4,4-Diethoxybutyl)-4-phenyl-butyramide (4). Isolated as an oil (100% yield). ¹H NMR (300 MHz, CDCl₃): δ 1.17 (t, 6H, J =6 Hz), 1.48-1.66 (m, 4H), 1.94 (quintet, 2H, J = 7 Hz), 2.13 (t, 2H, J = 7 Hz), 2.62 (t, 2H, J = 7 Hz), 3.22 (quartet, 2H, J =6 Hz), 3.39-3.52 (m, 2H), 3.56-3.68 (m, 2H), 4.45 (t, 1H, J =5.5 Hz), 5.82 (brs, 1H), 7.10–7.20 (m, 3H), 7.21–7.29 (m, 2H): ¹³C NMR and DEPT (75 MHz, CDCl₃) $\delta = 15.3$ (CH₃), 24.6 (CH₂), 27.2 (CH₂), 31.1 (CH₂), 35.2 (CH₂), 36.0 (CH₂), 39.2 (CH₂),61.5 (CH₂), 102.7 (CH), 125.9 (CH), 128.4 (CH), 128.5 (CH), 141.5 (C), 172.7 (C).

N-(4,4-Diethoxybutyl)-4-(4-methylphenyl)-butyramide (4a). Isolated as an oil (100%). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.19$ (t, 6H, J = 7 Hz), 1.53-1.70 (m, 4H), 1.93 (quin., 2H, J = 8 Hz),2.14 (t, 2H, J = 8 Hz), 2.31 (s, 3H), 2.60 (t, 2H, J = 8 Hz), 3.24(quartet, 2H, J = 6 Hz), 3.43–3.55 (m, 2H), 3.57–3.70 (m, 2H), 4.47 (t, 1H, J = 5.5 Hz), 5.62 (brs, 1H), 7.04 (d, 2H, J = 5.5 Hz), 7.08 (d, 2H, J = 5.5 Hz): ¹³C NMR and DEPT (75 MHz, CDCl₃) $\delta = 15.3 \text{ (CH}_3), 21.0 \text{ (CH}_3), 24.6 \text{ (CH}_2), 27.3 \text{ (CH}_2), 31.1 \text{ (CH}_2),$ 34.8 (CH₂), 36.0 (CH₂), 39.2 (CH₂), 61.5 (CH₂), 102.7 (CH), 128.4 (CH), 129.1 (CH), 135.4 (C), 138.4 (C), 172.7 (C).

N-(4,4-Diethoxybutyl)-4-(3,4-dimethoxy)phenyl-butyramide (4b). Isolated as an oil (100%). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.05-1.22$ (m, 6H), 1.45-1.66 (m, 4H), 1.81-1.99 (m, 2H), 2.06-2.19 (m, 2H), 2.46-2.62 (m, 2H), 3.15-3.30 (m, 2H), 3.35-3.50 (m, 2H), 3.51–3.66 (m, 2H), 3.77–3.88 (m, 6H), 4.36–3.49 (m, 1H), 5.63–5.91 (brs, 1H), 6.59–6.80 (m, 3H). ¹³C NMR and DEPT (75 MHz, CDCl₃) $\delta = 15.3$ (CH₃), 24.6 (CH₂), 27.4 (CH₂), 31.1 (CH₂), 34.8 (CH₂), 35.9 (CH₂), 39.2 (CH₂), 55.8 (CH₃), 55.9 (CH₃), 61.5 (CH₂), 102.7 (CH), 111.2 (CH), 111.8 (CH), 120.3 (CH), 134.2 (C), 147.2 (C), 148.8 (C), 172.7 (C).

N-(4,4-Diethoxybutyl)-4-(2,5-dimethoxy)phenyl-butyramide (4c). Isolated as an oil (100%). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.17$ (t, 6H, J = 7 Hz), 1.52–1.66 (m, 4H), 1.89 (quintet, 2H, J = 7.5 Hz, 2.16 (t, 2H, J = 7.5 Hz), 2.60 (t, 2H, J = 7.5 Hz), 3.24 (q, 2H, J = 6 Hz), 3.43-3.50 (m, 2H), 3.58-3.75 (m, 2H), 3.73 (s, 3H), 3.75 (s, 3H), 4.45 (t, 1H, J = 5.5 Hz), 5.75 (brs, 1H). ¹³C NMR and DEPT (75 MHz, CDCl₃) $\delta = 15.38$ (CH₃), 24.75 (CH₂), 25.96 (CH₂), 29.82 (CH₂), 31.14 (CH₂), 36.29 (CH₂), 39.27 (CH₂), 55.72 (CH₃), 56.00 (CH₃), 61.48 (CH₂), 102.74 (CH), 111.26 (CH), 111.38 (CH), 116.42 (CH), 131.26 (C), 151.80 (C), 153.57 (C), 172.93 (C).

N-(4,4-Diethoxybutyl)-4-(4-chlorophenyl)-butyramide Isolated as an oil (100%). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.18$ (t, 6H, J = 7 Hz), 1.45-1.66 (m, 4H), 1.81-1.99 (m, 2H), 2.12 (t, 4H)2H J = 7 Hz), 2.59 (t, 2H, J = 7 Hz), 3.25 (qt, 2H, J = 6.5 Hz), 3.38-3.53 (m, 2H), 3.56-3.68 (m, 2H), 4.44 (t, 1H, J = 7.5 Hz), 5.68 (brs, 1H), 7.08 (d, 2H, J = 8 Hz), 7.22 (d, 2H, J = 8 Hz). ¹³C NMR and DEPT (75 MHz, CDCl₃) $\delta = 15.3$ (CH₃), 24.6 (CH₂), 27.0 (CH₂), 31.1 (CH₂), 34.5 (CH₂), 35.7 (CH₂), 39.2 (CH₂), 61.5 (CH₂), 102.7 (CH), 128.4 (CH), 129.8 (CH), 131.6 (C), 140.0 (C), 172.4 (C).

N-(4,4-Diethoxybutyl)-4-(4-bromophenyl)-butyramide Isolated as an oil (100%). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.17$ (t, 6H, J = 7 Hz), 1.49-1.67 (m, 4H), 1.90 (quin, 2H, J = 7 Hz),

2.11 (t, 2H, J = 7 Hz), 2.57 (t, 2H, J = 7.5 Hz), 3.22 (quartet, 2H, J = 6.5 Hz), 3.39-3.52 (m, 2H), 3.56-3.69 (m, 2H), 4.45 (t, 2H)1H, J = 5 Hz), 5.91 (brs, 1H), 7.02 (d, 2H, J = 8 Hz), 7.36 (d, 2H, J = 8 Hz). ¹³C NMR and DEPT (75 MHz, CDCl₃) δ = 15.3 (CH₃), 24.6 (CH₂), 26.9 (CH₂), 31.1 (CH₂), 34.6 (CH₂), 35.7 (CH₂), 39.2 (CH₂), 61.5 (CH₂), 102.7 (CH), 119.7 (C), 130.2 (CH), 131.4 (CH), 140.5 (C), 172.4 (C).

N-(4,4-Diethoxybutyl)-4-(4-biphenyl)-butyramide (4f). Isolated as an oil (100%). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.12$ (t, 6H, J = 7 Hz), 1.45–1.66 (m, 4H), 1.99 (quintet, 2H, J = 7 Hz), 2.18 (t, 2H, J = 7 Hz), 2.68 (t, 2H J = 7 Hz), 3.25 (quartet, 2H, J = 6.5 Hz), 3.38-3.53 (m, 2H), 3.56-3.68 (m, 2H), 4.49 (t, 1H, J = 7.5 Hz), 5.83 (brs, 1H), 7.19–7.59 (m, 9H); ¹³C NMR and DEPT (75 MHz, CDCl₃) $\delta = 15.4$ (CH3), 24.6 (CH₂), 27.1 (CH₂), 31.1 (CH₂), 34.9 (CH₂), 36.0 (CH₂), 39.3 (CH₂), 61.5 (CH₂), 102.7 (CH), 127.0 (CH), 127.1 (CH), 128.7 (CH), 128.9 (CH), 138.9 (C), 140.7 (C), 141.0 (C), 172.7 (C).

N-(4,4-Diethoxybutyl)-4-(3-bromophenyl)-butyramide Isolated as an oil (100%). ¹H NMR (300 MHz, CDCl₃) $\delta = 1.18$ (t, 6H, J = 7 Hz), 1.49-1.67 (m, 4H), 1.92 (quin, 2H, J = 7 Hz),2.13 (t, 2H, J = 7 Hz), 2.60 (t, 2H, J = 7.5 Hz), 3.24 (quartet, 2H, J = 6.5 Hz), 3.39-3.52 (m, 2H), 3.56-3.69 (m, 2H), 4.46 (t, 2H)1H, J = 5 Hz), 5.69 (brs, 1H), 7.08-7.16 (m, 2H), 7.26-7.33 (m, 2H): 13 C NMR and DEPT (75 MHz, CDCl₃) $\delta = 15.3$ (CH₃), 24.6 (CH₂), 26.9 (CH₂), 31.1 (CH₂), 34.9 (CH₂), 35.7 (CH₂), 39.2 (CH₂), 61.5 (CH₂), 102.7 (CH), 122.4 (C), 127.2 (CH), 129.1 (CH), 130.0 (CH), 131.5 (CH), 143.9 (C), 172.3 (C).

N-(4,4-Diethoxybutyl)-4-(3-bromo-4-methoxyphenyl)-butyramide (4h). Isolated as an oil (100%). ¹H NMR (500 MHz, CDCl₃): $\delta = 1.16$ (t, 6H, J = 7 Hz), 1.51–1.65 (m, 4H), 1.88 (quin, 2H, J = 7 Hz, 2.11 (t, 2H, J = 7 Hz), 2.53 (t, 2H, J = 7.5 Hz), 3.24(quartet, 2H, J = 6.5 Hz), 3.40-3.49 (m, 2H), 3.55-3.64 (m, 2H), 3.83 (s, 3H), 4.45 (t, 1H, J = 5 Hz), 5.76 (brs, 1H), 6.78 (d, 1H, J = 58.5 Hz), 7.04 (dd, 1H, J = 8.5, 2.0 Hz), 7.32 (d, 1H, J = 2.0 Hz).: ¹³C NMR and DEPT (125 MHz, CDCl₃) δ = 15.38 (CH3), 24.62 (CH2), 27.23 (CH2), 31.13 (CH2), 34.00 (CH2), 35.80 (CH2), 39.28 (CH2), 56.33 (CH3), 61.56 (CH2), 102.73 (CH), 111.45 (C), 112.02 (CH), 128.46 (CH), 135.31 (CH), 135.31 (C), 154.17 (C), 172.58 (C).

N-(4,4-Diethoxybutyl)-4-(4-methoxyphenyl)-butyramide Isolated as an oil (100%) ¹H NMR (300 MHz, CDCl₃): $\delta = 1.17$ (t, 6H, J = 7 Hz), 1.45-1.66 (m, 4H), 1.89 (quintet, 2H, J = 7 Hz),2.12 (t, 2H J = 7 Hz), 2.55 (t, 2H, J = 7 Hz), 3.22 (quartet, 2H, J = 6 Hz), 3.38–3.53 (m, 2H), 3.56–3.68 (m, 2H), 3.75 (s, 3H), 4.45 (t, 1H, J = 5 Hz), 5.79 (brs, 1H), 6.79 (d, 2H, J = 8 Hz), 7.06(d, 2H, J = 8 Hz); ¹³C NMR and DEPT (75 MHz, CDCl₃) $\delta =$ 15.3 (CH₃), 24.6 (CH₂), 27.4 (CH₂), 31.1 (CH₂), 34.3 (CH₂), 35.9 (CH₂), 39.2 (CH₂), 55.3 (CH₃), 61.5 (CH₂), 102.7 (CH), 113.8 (CH), 129.4 (CH), 133.6 (C), 157.8 (C), 172.8 (C).

N-(4,4-Diethoxybutyl)-4,4-diphenyl-butyramide. Isolated as an oil (88%) ¹H NMR (300 MHz, CDCl₃): $\delta = 1.18$ (t, 6H, J =7 Hz), 1.45-1.66 (m, 4H), 2.07 (t, 2H, J = 7 Hz_), 2.39 (qu, 2H, J = 7 Hz), 3.21 (quartet, 2H, J = 6.5 Hz), 3.38–3.53 (m, 2H), 3.56-3.68 (m, 2H), 3.91 (t, 1H, J = 8 Hz), 4.46 (t, 1H, J =7.5 Hz), 5.57 (brs, 1H), 7.10–7.32 (m, 10H). ¹³C NMR and DEPT (75 MHz, CDCl₃) $\delta = 15.3$ (CH₃), 24.6 (CH), 31.1 (CH₂), 31.2

(CH₂), 35.0 (CH₂), 39.2 (CH₂), 50.6 (CH), 61.4 (CH₂), 102.7 (CH), 126.3 (CH), 127.9 (CH), 128.5 (CH), 144.3 (C), 172.4 (C).

5-Phenyl-pentanoic acid (3,3-diethoxybutyl)-amide (9). Isolated as an oil (95%): ¹H NMR (300 MHz, CDCl₃): $\delta = 1.18$ (t, 6H, J = 7 Hz), 1.50-1.73 (m, 8 H), 2.15 (t, 2H, J = 7 Hz), 2.61(t, 2H, J = 7 Hz), 3.23 (q, 2H, J = 6 Hz), 3.40-3.53 (m, 2H), 3.55-3.69 (m, 2H), 4.46 (t, 1H, J = 5 Hz), 5.71 (brs, 1H), 7.12-7.19 (m, 2H)3H), 7.22–7.27 (m, 2H). ¹³C NMR and DEPT (75 MHz, CDCl₃) $\delta = 15.3 \, (CH_3), 24.6 \, (CH_2), 25.4 \, (CH_2), 31.1 \, (CH_2), 35.7 \, (CH_2),$ 36.7 (CH₂), 39.2 (CH₂), 61.5 (CH₂), 102.7 (CH), 125.7 (CH), 128.3 (CH), 128.4 (CH), 142.2 (C), 172.8 (C).

N-(5,5-Diethoxypentyl)-4-phenyl-butyramide (10). Isolated as an oil (100%) ¹H NMR (300 MHz, CDCl₃): $\delta = 1.18$ (t, 6H, J = 7 Hz), 1.32-1.67 (m, 6H), 1.95 (quintet, 2H, J = 7.4 Hz), 2.15 (t, 2H, J = 7.4 Hz, 2.64 (t, 2H, J = 7.4 Hz), 3.22 (q, 2H, J = 6 Hz), 3.40-3.55 (m, 2H), 3.56-3.71 (m, 2H), 4.45 (t, 1H, J = 5.6 Hz), 5.50(brs, 1H), 7.13-7.23 (m, 3H), 7.24-7.31 (m, 2H). ¹³C NMR and DEPT (75 MHz, CDCl₃) $\delta = 15.3$ (CH₃), 22.1 (CH₂), 27.2 (CH₂), 29.5 (CH₂), 33.3 (CH₂), 35.2 (CH₂), 36.0 (CH₂), 39.4 (CH₂), 61.1 (CH₂), 102.8 (CH), 125.9 (CH), 128.4 (CH), 128.5 (CH), 141.5 (C), 172.6 (C).

2-Benzyl-N-(4,4-diethoxybutyl)benzamide (11a). Isolated as an oil (100%) ¹H NMR (300 MHz, CDCl₃): $\delta = 1.16$ (t, 6H, J = 7 Hz), 1.45-1.65 (m, 4H), 3.30 (q, 2H, J = 6 Hz), 3.35-3.50 (m, 2H), 3.53-3.68 (m, 2H), 4.17 (s, 2H), 4.43 (t, 1H, J = 5.3 Hz), 5.90 (brs, 1H), 7.07–7.37 (m, 9H). ¹³C NMR and DEPT (75 MHz, $CDCl_3$) $\delta = 15.3 (CH_3), 24.4 (CH_2), 31.1 (CH_2), 38.9 (CH_2), 39.6$ (CH₂), 61.4 (CH₂), 102.6 (CH), 126.0 (CH), 126.3 (CH), 127.1 (CH), 128.4 (CH), 129.0 (CH), 130.9 (CH), 137.0 (C), 138.8 (C), 140.9 (C), 170.0 (C).

N-(4,4-Diethoxybutyl)-2-phenoxybenzamide (11b). Isolated as an oil (95%). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.13$ (t, 3H, J = 7 Hz), 1.14 (t, 3H, J = 7 Hz), 1.53-1.67 (m, 4H), 3.30-3.61 (m, 6H), 4.41-4.40 (m, 1H), 6.80 (dd, 1H, J = 8.2, 1.7 Hz), 7.00-7.04(m, 2H), 7.13–7.20 (m, 2H), 7.29–7.41 (m, 3H), 7.69 (brs, 1H), 8.20 (dt, 1H, J = 7.8, 1.5 Hz). ¹³C NMR and DEPT (75 MHz, CDCl₃) $\delta = 15.3 \text{ (CH}_3), 24.7 \text{ (CH}_2), 30.9 \text{ (CH}_2), 39.5 \text{ (CH}_2), 61.1 \text{ (CH}_2),$ 102.5 (CH), 118.4 (CH), 119.4 (CH), 123.7 (CH), 124.3 (C), 124.6 (CH), 130.1 (CH), 132.2 (CH), 132.5 (CH), 155.3 (C), 155.6 (C), 164.7 (C).

General procedure for the synthesis of the 2-hydroxypyrrolidino-lactams 7d-h

A solution of the 4,4-diethoxybutyl butyramide (3 mmol) in acetone (50 ml) and 1 M HCl (12 ml) was allowed to stand at room temperature for 30 min., by which time TLC showed no starting material remaining. The reaction mixture was basified with 1 M NaHCO₃ solution (30 ml) and the acetone removed under reduced pressure on a rotary evaporator. The product was extracted from the aqueous residue with CH_2Cl_2 (3 × 50 ml). The combined organic extracts were dried (K₂CO₃), filtered and evaporated to give the 2-hydroxypyrrolidino-lactam, used without further purification. NMR analysis showed them to be a complex mixture of ~20% aldehyde and amide rotomers.

General procedure for the triflic acid-mediated cyclisation

A solution of the acetal (5 mmol) in chloroform (10 mL) was added over 10 min to a heated (65 °C), stirred mixture of triflic acid (50 mmol) in chloroform (40 mL). The reaction was heated under gentle reflux for a given period. On cooling to ambient temperatures, water (20 ml) was added. The reaction mixture was transferred to a separating funnel and the lower layer separated. The aqueous layer was extracted with DCM (50 ml) and the combined organic extracts dried (K₂CO₃). Filtration and evaporation in vacuo gave the crude products that were separated by column chromatography on silica.

7-Phenyl-2,3,6,10b-tetrahydro-1*H*-pyrrolo[2,1-*a*]isoquinolin-5one (2). Reaction heated to reflux for 3 h, purified by elution with 1:1 CH₂Cl₂/Et₂O to 2%MeOH/Et₂O and fraction 2 isolated as a solid from trituration with Et₂O (64%): mpt 157–9 °C. HRMS Theoretical Mass: 263.12551, found: 263.12589. ¹H NMR (300 MHz, CDCl₃): $\delta = 1.91-2.25$ (m, 3H), 2.61-2.79 (m, 1H), 3.41-3.55 (m, 2H), 3.63 (d, 1H, J = 18 Hz), 3.66-3.77 (m, 1H), 4.63-4.77 (m, 1H), 7.15–7.45 (m, 8H). ¹³C NMR and DEPT (75 MHz, CDCl₃): $\delta = 23.4 \text{ (CH}_2), 31.7 \text{ (CH}_2), 37.0 \text{ (CH}_2), 44.8 \text{ (CH}_2), 59.7 \text{ (CH)},$ 123.2 (CH), 126.6 (CH), 127.4 (CH), 128.4 (CH), 129.1 (CH), 129.2 (CH), 130.8 (C), 136.8 (C), 140.2 (C), 140.9 (C), 167.7 (C).

1,2,3,6,10b-Pentahydro-1H-pyrrolo[2,1-a]-dibenzo[c,e]azocin-5one (3). Reaction heated to reflux for 3 h, purified by elution with 1:1 CH₂Cl₂/Et₂O to 2%MeOH/Et₂O and as fraction 1 isolated as an oil (29%). HRMS Theoretical Mass: 263.12551: found: 263.12567. ¹H NMR (300 MHz, CDCl₃): $\delta = 1.66-1.86$ (m, 1H), 1.98-2.20 (m, 2H), 2.35-2.50 (m, 1H), 3.12 (d, 1H, J = 15 Hz), 3.57 (d, 1H, J = 15 Hz), 3.60-3.80 (m, 2H), 4.27 (t, 1H, J = 8 Hz), 7.25–7.55 (m, 8H). 13 C NMR and DEPT (75 MHz, CDCl₃): $\delta =$ 23.4 (CH₂), 33.6 (CH₂), 43.3 (CH₂), 48.4 (CH₂), 53.5 (CH₂), 56.7 (CH), 126.0 (CH), 127.5 (CH), 127.7 (CH), 127.9 (CH), 128.3 (CH), 128.5 (CH), 128.6 (CH), 130.0 (CH), 132.6 (C), 137.1 (C), 141.2 (C), 142.6 (C), 168.7 (C).

6-Aza-tricyclo[9.4.0.0*2,6*]pentadeca-1(11),12,14-trien-7-one (5). Reaction heated to reflux for 4 h, purified by elution with Et₂O-2% MeOH/Et₂O, isolated as an oil (70% yield). HRMS theoretical 215.13047, found 215.13000: FTIR (film) 2947, 2873, 1624, 1455, 1420, 754 cm⁻¹.

14-Methyl-6-aza-tricyclo[9.4.0.0*2,6*]pentadeca-1(11),12,14trien-7-one (5a). Reaction heated to reflux for 4 h, purified by elution with Et₂O-2% MeOH/Et₂O, isolated as an oil (70% yield). HRMS Theoretical Mass: 229.14611, found 229.14545.

13,14-Dimethoxy-6-aza-tricyclo[9.4.0.0*2,6*]pentadeca-1(11), 12,14-trien-7-one (5b). From the acetal at reflux for 1 h, purified by elution with Et₂O-2% MeOH/Et₂O, isolated as a white solid, recrystallised from EtOAc/petrol (80% yield), m. pt. 116-7 °C. HRMS Theoretical Mass: 275.15160, found 275.15154.

12,15-Dimethoxy-6-aza-tricyclo[9.4.0.0*2,6*]pentadeca-1(11), 12,14-trien-7-one (5c). Isolated from the acetal at room temperature for 21 h, purified by elution with Et₂O-2% MeOH/Et₂O, isolated as a white solid, recrystallized from EtOAc/petrol (26% yield), m. pt. 126-7 °C. HRMS Theoretical Mass: 275.15160, found 275.15102. FTIR (solid) 1634, 1471, 1417, 1252, 1141, 1090, 784, 718 cm⁻¹.

14-Chloro-6-aza-tricyclo[9.4.0.0*2,6*|pentadeca-1(11),12,14trien-7-one (5d). Prepared from the 2-hydroxypyrrolidine, heated to reflux for 18 h, purified by elution with Et₂O-2% MeOH/Et₂O, isolated as an oil (59% yield). HRMS: Theoretical Mass: 249.09149, found 249.09081.

14-Bromo-6-aza-tricyclo[9.4.0.0*2,6*]pentadeca-1(11),12,14**trien-7-one (5e).** Prepared from the 2-hydroxypyrrolidine, heated to reflux for 18 h, purified by elution with Et₂O-2% MeOH/Et₂O, isolated as an oil (55% yield). HRMS Theoretical Mass: 293.04098, found 293.03969.

14-Phenyl-6-aza-tricyclo[9.4.0.0*2,6*]pentadeca-1(11),12,14**trien-7-one (5f).** Prepared from the 2-hydroxypyrrolidine, heated to reflux for 18 h, purified by elution with Et₂O-2% MeOH/Et₂O, isolated as an oil (45% yield). HRMS Theoretical Mass: 291.16177; found: 291.16191.

13-Bromo-6-aza-tricyclo[9.4.0.0*2,6*]pentadeca-1(11),12,14**trien-7-one (5g).** From the 2-hydroxypyrrolidine, heated to reflux for 18 h, purified by elution with Et₂O-4% MeOH/Et₂O, isolated as white solid (Et₂O/petrol) (60% yield). HRMS Theoretical Mass: 293.04098: found 293.04037. 1H NMR (500 MHz, CDCl₃) at 332 K: $\delta = 1.68-1.80$ (brm, H-9), 1.87–2.11 (m, H-3,4,4,9), 2.26– 2.35 (brm, H-8), 2.40–2.52 (m, H-3, 8), 2.61–2.70 (brm, H-10), $2.96 \text{ (ddd, H-10, J} = 3.8, 11.9, 13.8 Hz), } 3.40-3.50 \text{ (brm, H-5)},$ 3.90 (ddd, H-5, J = 4.1, 8.6, 12.4 Hz), 5.01 (t, H-2, J = 7 Hz),7.02 (d, H-15, J = 8.3 Hz), 7.25 (d, H-12, J = 2.2 Hz), 7.32 (dd, H-14, J=2.2, 8.3 Hz). ^{13}C NMR and DEPT (150 MHz, CDCl₃) at 332 K: $\delta = 22.60, 26.99, 32.22, 33.48, 36.79, 45.88, 62.34$ (2-C), 120.99 (C-13), 128.54 (C-15), 129.50 (C-14), 134.01 (C-12), 139.54 (C-1), 140.27 (C-11), 172.14 (C-7).

13-Bromo-14-methoxy-6-aza-tricyclo[9.4.0.0*2,6*]pentadeca-1(11),12,14-trien-7-one (5h). Isolated from the acetal at reflux for 1 h, purified by elution with Et₂O-2% MeOH/Et₂O, isolated as a white solid, recrystallized from EtOAc/petrol (80% yield), m. pt. 116-7 °C. HRMS Theoretical Mass: 323.05154, found 323.05138. ¹H NMR (600 MHz, CDCl₃) at 331 K: $\delta = 1.64-1.77$ (brm, 1H), 1.87–2.03 (m, 3H), 2.04–2.13 (m, 1H), 2.26–2.34 (brm, 1H), 2.41– 2.52 (m, 2H), 2.54-2.2 (brm, 1H), 2.89 (ddd, 1H, J = 3.8, 12.1, 14 Hz), 3.46 (dt, 1H, J = 8.0, 12.0 Hz), 3.86 (s, 3H), 3.89 (ddd, 1H-5, J = 12.1, 8.6, 4.2 Hz), 5.00 (t, 1H, J = 7.1 Hz), 6.66 (s, 1H), 7.26 (s, 1H). ¹³C NMR and DEPT (150 MHz, CDCl₃) at 333 K: $\delta = 22.86 \text{ (CH}_2), 27.62 \text{ (CH}_2), 31.24 \text{ (CH}_2), 33.43 \text{ (CH}_2), 46.30$ (CH₂), 56.61 (CH₃), 63.01 (CH), 110.68 (C), 111.19 (CH), 131.89 (C), 136.06 (CH), 141.04 (C), 154.65 (C), 172.69 (C).

6-Aza-tricyclo[9.4.0.0*2,6*|pentadeca-1(15),11,13-triene (6). A solution of 6-aza-tricyclo[9.4.0.0*2,6*]pentadeca-1(11),12,14trien-7-one (0.9 g, 4.2 mmol) in dry THF (5 ml) was added, dropwise, to a stirred solution of 1 M LiAlH₄ (4.2 ml, 4.2 mmol) in dry THF (20 ml) and the reaction mixture heated to reflux for 2 h. On cooling to 0 °C, 2 M NaOH (0.6 ml) was carefully added, drop-wise, the reaction mixture stirred for 30 min. and then Et₂O (50 ml) was added. The white solid was removed, washed with Et₂O $(2 \times 50 \text{ ml})$ and the combined organics dried (K_2CO_3), filtered and concentrated in vacuo to give the title compound as an oil (0.8 g, 88% yield). HRMS Theoretical Mass: 201.15120, found 201.15131. Attempts to form an HCl salt resulted in a sticky,

hygroscopic gum. A sample was treated with 1 equivalent of picric acid (35% water) in ethanol to give the picrate, (m pt 162–4 °C).

(2R,10R)(2S,10S)-10-Phenyl-6-aza-tricyclo[9.4.0.0*2,6*] pentadeca-1(15),11,13-triene-7-one (8). From the acetal at reflux for 3 h, purified by elution with Et₂O and the solid triturated with petrol (73%), a small sample was re-crystallized from CH₂Cl₂/petrol, m. pt. 162–4 °C. ¹H NMR (500 MHz, CDCl₃) at 332 K: $\delta = 1.75 - 2.60$ (m, 7H), 3.60 - 3.79 (m, 1H), 3.87 (dt, 1H, J = 9.5, 2.4 Hz), 4.53 (dd, 1H, J = 12.3, 3.2 Hz), 4.93 (dd, 1H, J = 10.9, 5.1 Hz, 6.71 (d, 1H, J = 6.8 Hz), <math>7.05-7.45 (m, 8H). ¹³C NMR and DEPT (125 MHz, CDCl₃) at 332 K: $\delta = 22.4$ (CH₂), 31.2 (CH₂), 32.7 (CH₂), 39.5 (CH₂), 43.3 (CH), 46.9 (CH₂), 65.8 (CH), 126.5 (CH), 126.6 (CH), 127.7 (CH), 127.9 (CH), 128.5 (CH), 128.7 (CH), 129.2 (CH), 138.9 (C), 141.0 (C), 142.9 (C), 172.3 (C).

4b,5,6,7-Tetrahydro-14H-dibenzo[c,f]pyrrolo[1,2-a]azocin-9-one (12a). From the acetal at reflux for 3 h, purified by elution with CH₂Cl₂-20% Et₂O/CH₂Cl₂, isolated as a white solid, recrystallised from EtOAc/petrol (95% yield), m. pt. 127-9 °C. HRMS Theoretical Mass: 263.13046, found 263. 12983. ¹H NMR (300 MHz, CDCl₃): $\delta = 1.80-2.17$ (m, 3H), 2.55-2.71 (m, 1H), 3.23-3.30 (m, 1H), 3.79 (d, 1H J = 14.5 Hz), 3.99-4.13(m, 1H), 4.3 (d, 1H, J = 14.5 Hz), 4.67 (d, 1H, J = 5.5 Hz), 7.12–7.41 (m, 7H), 7.52 (dd, 1H, J = 7, 1 Hz). ¹³C NMR and DEPT (75 MHz, CDCl₃): $\delta = 23.0$ (CH₂), 29.0 (CH₂), 41.0 (CH₂), 44.4 (CH₂), 60.6 (CH), 125.2 (CH), 125.6 (CH), 127.1 (CH), 127.2 (CH), 127.9 (CH), 129.6 (CH), 130.6 (CH), 131.2 (CH), 134.9 (C), 136.4 (C), 138.0 (C), 139.2 (C), 171.0 (C).

4b,5,6,7-Tetrahydro-14-oxa-dibenzo[c,f]pyrrolo[1,2-a]azocin-9one (12b). From the acetal at reflux for 2 h, purified by elution with CH₂Cl₂-10% Et₂O/CH₂Cl₂, (66% yield), m. pt. 144-6 °C. HRMS Theoretical Mass: 265.10973, found 265.10973. ¹H NMR (600 MHz, CDCl₃): $\delta = 1.85-1.96$ (m, 6–1H), 1.96–2.04 (m, 5-1H), 2.04-2.10 (m, 6-1H), 2.62 (dd, 5-1H, J = 5.3, 12.8 Hz), 3.40-3.48 (m, 7-1H), 3.87-3.95 (m, 7-1H), 4.72 (d, 4d-1H, J =6.8 Hz), 7.10-7.16 (m, 2H), 7.22 (d, 1H, J = 7.9 Hz), 7.27 (t, 1H, J = 7.9 Hz, 7.32-7.48 (m, 2H), 7.42 (dt, 1H, J = 7.9, 1.2 Hz), 7.58 (dd, 1H, J = 7.6, 1.2 Hz). ¹³C NMR and DEPT (150 MHz, CDCl₃): $\delta = 23.18$ (6-CH₂), 28.75 (5-CH₂), 45.17 (7-CH₂), 58.79 (4b-CH), 122.57 (CH), 122.74 (CH), 124.71 (CH), 125.99 (CH), 126.29 (CH), 127.52 (CH), 129.22 (C), 129.50 (CH), 132.23 (CH), 132.61 (C), 152.82 (C), 157.63 (C), 168.08 (C).

(4,4-Diethoxy-butyl)-(4-phenyl-butyl)-amine (13). A solution of the amide 4 (2.0 g, 6.5 mmol) in dry THF (20 ml) was added to a stirred solution of 1 M LAH (30 ml, 30 mmol) in THF (30 ml) under argon at 0 °C and the reaction was heat under gentle reflux for 24 h. The reaction mixture was cooled to 0 °C and H₂O (1 ml), then 2 M NaOH (2 ml) were carefully add and the reaction mixture stirred until a white solid formed. Et₂O (50 ml) was then addded, the solid collected which was thoroughly washed with Et₂O (4 \times 25 ml). The combined organic filtrate and washings were dried (K₂CO₃), filter and concentrate in vacuo to give a colourless oil: 1.9 g (~100%). ¹H NMR (500 MHz, CDCl₃): $\delta = 1.19$ (t, 6H, J = 7 Hz), 1.50–1.57 (m, 4H), 1.61–1.70 (m, 4H), 2.58–2.65 (m, 6H), 3.45-3.54 (m, 2H), 3.59-3.67 (m, 2H), 4.48 (t, 1H, J = 5.7 Hz), 7.14–7.18 (m, 3H), 7.24–7.28 (m, 2H); ¹³C NMR and DEPT (125 MHz, CDCl₃): $\delta = 15.41$ (CH₃), 25.51 (CH₂), 29.32 (CH₂), 29.83 (CH₂), 31.54 (CH₂), 35.92 (CH₂), 49.89 (CH₂), 49.94 (CH₂), 61.06 (CH₂), 61.16 (CH₂), 102.90 (CH), 125.74 (CH), 128.40 (CH), 128.53 (CH), 142.56 (CH).

Crystal data and structure refinement

12,15-Dimethoxy-6-aza-tricyclo[9.4.0.0*2,6*|pentadeca-1(11), **12,14-trien-7-one (5c).** Chemical formula $C_{16}H_{21}NO_3$; Formula weight 275.34; Temperature 150(2) K; Radiation, wavelength $MoK\alpha$, 0.71073 Å; Crystal system, space group monoclinic, $P2_1/c$; Unit cell parameters $a = 8.3263(9) \text{ Å}; \alpha = 90^{\circ}; b = 18.599(2) \text{ Å}; \beta =$ $107.795(2)^{\circ}$; c = 9.2541(10) Å; $\gamma = 90^{\circ}$; Cell volume 1364.5(3) Å³ Z 4; Calculated density 1.340 g/cm³; Absorption coefficient μ 0.092 mm^{-1} ; F(000) 592; Crystal colour and size colourless, $0.40 \times$ 0.12 × 0.02 mm³; Data collection method Bruker SMART APEX CCD diffractometer ω rotation with narrow frames; θ range for data collection 2.56 to 28.27°; Index ranges h -10 to 11, k -24 to 24, 1 -12 to 12; Completeness to $\theta = 26.00^{\circ} 99.4\%$; Reflections collected 11463; Independent reflections 3261 (R_{int} = 0.0276); Reflections with $F^2>2\sigma$ 2760; Absorption correction semi-empirical from equivalents; Min. and max. transmission 0.9641 and 0.9982; Structure solution direct methods; Refinement method Full-matrix least-squares on F²; Weighting parameters a, b 0.0681, 0.3291; Data/restraints/parameters 3261/0/181; Final R indices $[F^2>2\sigma]$ R1 = 0.0439, wR2 = 0.1168; R indices (all data) R1 = 0.0519, wR2 = 0.1221; Goodness-of-fit on F² 1.066; Largest and mean shift/su 0.000 and 0.000; Largest diff, peak and hole 0.343 and -0.260 e Å⁻³.

2R,10R)(2S,10S)-10-Phenyl-6-aza-tricyclo[9.4.0.0*2,6*]-pentadeca-1(15),11,13-triene-7-one (8). Chemical formula $C_{20}H_{21}NO$; Formula weight 291.38; Temperature 150(2) K; Radiation, wavelength MoKα, 0.71073 Å; Crystal system, space group monoclinic, $P2_1/c$; Unit cell parameters $a = 7.6569(8) \text{ Å } \alpha = 90^\circ, b =$ 9.6582(11) Å $\beta = 90.423(2)^{\circ}$, c = 20.404(2) Å $\gamma = 90^{\circ}$; Cell volume 1508.9(3) Å³; Z 4; Calculated density 1.283 g/cm³; Absorption coefficient μ 0.078 mm⁻¹; F(000) 624; Crystal colour and size colourless, $0.48 \times 0.46 \times 0.43$ mm³; Data collection method Bruker SMART APEX CCD diffractometer ω rotation with narrow frames; θ range for data collection 3.34 to 28.29°; Index ranges h -10 to 10, k -12 to 12, 1 -26 to 26; Completeness to $\theta = 26.00^{\circ}$ 99.3%; Reflections collected 12268; Independent reflections 3580 $(R_{int} = 0.0327)$; Reflections with $F^2 > 2\sigma$ 3271; Absorption correction semi-empirical from equivalents; Min. and max. transmission 0.9634 and 0.9671; Structure solution direct methods; Refinement method Full-matrix least-squares on F2; Weighting parameters a, b 0.0720, 0.4836; Data/restraints/parameters 3580/0/200; Final R indices $[F^2>2\sigma]$ R1 = 0.0440, wR2 = 0.1201; R indices (all data) R1 = 0.0473, wR2 = 0.1231; Goodness-of-fit on F^2 1.041; Extinction coefficient 0.027(4); Largest and mean shift/su 0.000 and 0.000; Largest diff. peak and hole 0.402 and -0.248 e Å⁻³.

4b,5,6,7-Tetrahydro-14*H***-dibenzo**[*c,f*]**pyrrolo**[1,2-*a*]**azocin-9-one (12a).** Chemical formula $C_{18}H_{17}NO$; Formula weight 263.33; Temperature 150(2) K; Radiation, wavelength MoKα, 0.71073 Å; Crystal system, space group triclinic, P \bar{l} ; Unit cell parameters a = 9.3106(8) Å α = 75.9490(10)° b = 10.4150(9) Å β = 88.8050(10)° c = 16.0747(14) Å γ = 63.5860(10)°; Cell volume 1347.4(2) ų Z 4; Calculated density 1.298 g/cm³; Absorption coefficient μ 0.080 mm $^{-1}$; F(000) 560; Crystal colour and size colourless, 0.32 ×

 0.28×0.18 mm³; Data collection method Bruker SMART APEX CCD diffractometer ω rotation with narrow frames θ range for data collection 2.63 to 28.30°; Index rangesh –12 to 12, k –13 to 13, 1 –20 to 21; Completeness to $\theta=26.00^\circ$ 97.7%; Reflections collected 11524; Independent reflections 6146 ($R_{int}=0.0221$); Reflections with $F^2>2\sigma$ 5332; Absorption correction semi-empirical from equivalents; Min. and max. transmission 0.9748 and 0.9857; Structure solution direct methods; Refinement method Full-matrix least-squares on F^2 ; Weighting parameters a, b 0.1039, 0.3159; Data/restraints/parameters 6146/0/361; Final R indices [$F^2>2\sigma$] R1 = 0.0520, wR2 = 0.1554; R indices (all data) R1 = 0.0583, wR2 = 0.1641. Goodness-of-fit on F^2 1.079; Largest and mean shift/su 0.000 and 0.000; Largest diff. peak and hole 0.476 and -0.433 e Å $^{-3}$.

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References

- 1 D. A. Horton, G. T. Bourne and M. L. Smythe, *Chem. Rev.*, 2003, **103**, 893, and references quoted therein.
- 2 E. J. Ariens, A. J. Beld, J. F. Rodrigues de Miranda and A. M. Simonis, in *The Receptors A Comprehensive Treatise*, R. D. O'Brien, Ed.; Plenum Press, New York, 1979, p 33.
- 3 P. R. Andrews and E. J. Lloyd, Med. Res. Rev., 1982, 2, 355.
- 4 E. S. Vermeulen, M. van Smeden, A. W. Schmidt, J. S. Sprouse, H. V. Wikström and C. J. Grol, *J. Med. Chem.*, 2004, 47, 5451.
- 5 M. Dukat, M. Taroua, A. Dahdouh, U. Siripurapu, D. Imad, B. R. Martin and R. A. Glennon, *Bioorg. Med. Chem. Lett.*, 2004, 14, 3651.
- 6 J. M. Keith, L. A. Gomez, R. L. Wolin, A. J. Barbier, S. J. Wilson, J. D. Boggs, C. Mazur, I. C. Fraser, B. Lord, L. Aluisio, T. W. Lovenberg and N. I. Carruthers, *Bioorg. Med. Chem. Lett.*, 2007, 17, 702.
- 7 C. Ma, S.-J. Liun, L. Xin, Q. Zhang, K. Ding, J. R. Falck and D.-S. Shin, *Chem. Letts.*, 2006, 35, 1010.
- 8 J. W. Lane, A. Estevezc, K. Mortarac, O. Callanc, J. R. Spencer and R. M. Williams, *Bioorg. Med. Chem. Lett.*, 2006, **16**, 3180.
- 9 N. E. Austin, K. Y. Avenell, I. Boyfield, C. L. Branch, M. C. Coldwell, M. S. Hadley, P. Jeffrey, A. Johns, C. N. Johnson, D. J. Nash, G. J. Riley, S. A. Smith, R. C. Stacey, G. Stemp, K. M. Thewlis and A. K. Vong, *Bioorg. Med. Chem. Lett.*, 1999, 9, 179.
- 10 W. L. Huang, X. Q. Song, S. X. Peng and Z. Y. Huang, *Yaoxue Xuebao*, 1990, 25, 815.
- 11 A. J. Bojarski, M. J. Mokrosz, S. C. Mino, A. Koziol, A. Wesolowska, E. Tatarczynska, A. Klodzinska and E. Chojnacka-Wojcik, *Bioorg. Med. Chem.*, 2002, 10, 87.
- 12 B. E. Maryanoff, D. F. McComsey, J. F. Gardocki, R. P. Shank, M. J. Costanzo, S. O. Nortey, C. R. Schneider and P. E. Setler, *J. Med. Chem.*, 1987, **30**, 1433.
- 13 G. Stemp, T. Ashmeade, C. L. Branch, M. S. Hadley, A. J. Hunter, C. N. Johnson, D. J. Nash, K. M. Thewlis, A. K. Vong, N. E. Austin, P. Jeffrey, K. Y. Avenell, I. Boyfield, J. J. Hagan, D. N. Middlemiss, C. Reavill, G. J. Riley, C. Routledge and M. Wood, *J. Med. Chem.*, 2000, 43, 1878.
- 14 F. D. King, Tetrahedron, 2007, 63, 2053.
- 15 L. Gang, R. Franzen, Q. Zhang and Y. Xu, Tetrahedron Lett., 2005, 46, 4255
- 16 B. E. Maryanoff, H.-C. Zhang, J. H. Cohen, I. J. Turchi and C. A. Maryanoff, *Chem. Rev.*, 2004, **104**, 1431.
- 17 G. Hilt, F. Galbiati and K. Harms, Synthesis, 2006, 3575.
- 18 Antidepressants: Past, Present and Future, in Handbook of Experimental Pharmacology, S. Preskorn, C. Y. Stanga, J. P. Feighner, and R. Ross, (eds.) 2004, 157, Springer.
- 19 Z. F. Solomko and A. N. Kost, Chem. Het. Cpds., 1975, 11, 1231.
- 20 G. Flynn, E. Giroux and R. Dage, J. Am. Chem. Soc., 1987, 109, 7914.
- 21 G. L. Grunwald, V. H. Dahanukar, P. Ching and K. R. Criscione, J. Med. Chem., 1996, 39, 3539.

- 22 H. Fuwa, Y. Okamura, Y. Morohashi, T. Tomita, T. Iwatsubo, T. Kan, T. Fukuyama and H. Natsugari, Tetrahedron Lett., 2004, 45, 2323.
- 23 S. Bergemann, R. Brecht, F. Büttner, D. Guénard, R. Gust, G. Seitz, M. T. Stubbs and S. Thoret, Bioorg. Med. Chem., 2003, 11, 1269.
- 24 S. Seto, A. Tanioka, M. Ikeda and S. Izawa, Bioorg. Med. Chem., 2005, **13**, 5717.
- 25 D. H. Jones, G. F. Stephenson, G. W. Spray and W. R. Wragg, J. Chem. Soc. C, 1969, 2176.
- 26 L. A. Paquette, L. B. Anderson, J. F. Hansen, S. A. Lang, Jr. and H. Berk, J. Am. Chem. Soc., 1972, 94, 4907.
- 27 A. M. Monro, R. M. Quinton and T. I. Wrigley, J. Org. Chem., 1963,
- 28 M. Qadir, J. Cobb, P. W. Sheldrake, N. Whittall, A. J. P. White, K. K. Hii, P. N. Horton and M. B. Hursthouse, J. Org. Chem., 2005, 70, 1552.
- 29 J.-L. Panayides, R. Pathak, H. Panagiotopoulos, H. Davids, M. A. Fernandes, C. B. de Koning and W. A. L. van Otterlo, Tetrahedron, 2007, 63, 4737.
- 30 O. O. Orazi, R. A. Corral and H. Giacco, J. Chem. Soc., Perkin Trans. 1, 1986, 1977.
- 31 R. A. Corral, Monograf. Acad, Nac. Cien. Exact. Fis. Nat., 1988, 3,

- 32 J. Zinczuk, I. H. Sorokin, O. Orfeo and R. A. Corral, J. Het. Chem., 1992, 29, 859.
- 33 M. Mizukami, H. Saito, T. Higuchi, M. Imai, H. Bando, N. Kawahara and S. Nagumo, Tetrahedron Lett., 2007, 48, 7228.
- 34 Y. Zhang, D. J. DeSchepper, T. M. Gilbert, K. K. S. Sai and D. A. Klumpp, Chem. Commun., 2007, 4032.
- 35 J. Jeener, B. H. Meier, P. Bachmann and R. R. Ernst, J. Chem. Phys., 1979, 71, 4546.
- 36 A. E. Aliev and D. Courtier-Murias, J. Phys. Chem. B, 2007, 111, 14034.
- 37 C. A. G. Haasnoot, F. A. A. M. DeLeeuw, H. P. M. DeLeew and C. Altona, Biopolymers, 1981, 20, 1211.
- 38 K. D. Moeller, P. W. Wang, S. Tarazi, M. R. Marzabadi and P. L. Wong, J. Org. Chem., 1991, 56, 1058.
- 39 H. E. Zaugg, Synthesis, 1984, 85.
- 40 D. Stephenson and G. J. Binsch, Magn. Reson., 1980, 37, 395; G. Hägele, M. Engelhardt, and M. Boenigk, Simulation und automatisierte Analyse von Kernresonanzspektren, VCH, Weinheim, 1987; gNMR, Version 5.0.6, NMR Simulation Program, Budzelaar PHM, 2006.
- 41 H. Shanan-Atidi and K. H. Bar-Eli, J. Phys. Chem., 1970, 74, 961.
- 42 M. J. Chapdeleine, and D. McLaren, GB 2251616A (1992) CA 117, 212302.